

## Introduction

### Controls of transplants (out of clinical trials):

- Screening for pathogens in stool
- Quantity (minimum 30g for 25g and 55g for 50g of stool)
- Macroscopic examination (consistency, presence of foreign elements, blood, etc.)

### No characterization of:

- Type of microorganisms (MO) present?
- Number of MO in a transplant?
- Percentage of bacterial viability?
- Proportion of other components of the fecal microbiota?

### Flow cytometry would allow:

- Enumeration of MO
- Proportion of viable MO: fluorescent staining with differing permeability based on membrane integrity



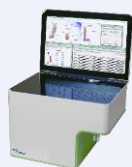
Native stool



Fecal suspension  
(0.9% NaCl +  
Glycerol)

## Materials et methods

### Tests on 2 cytometers

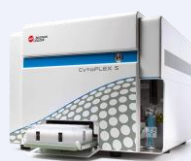


CyFlow® Cube 6  
(Sysmex®)

Staining: CyStain™  
Green/Red

Triplicate, 23 stools

Blue laser



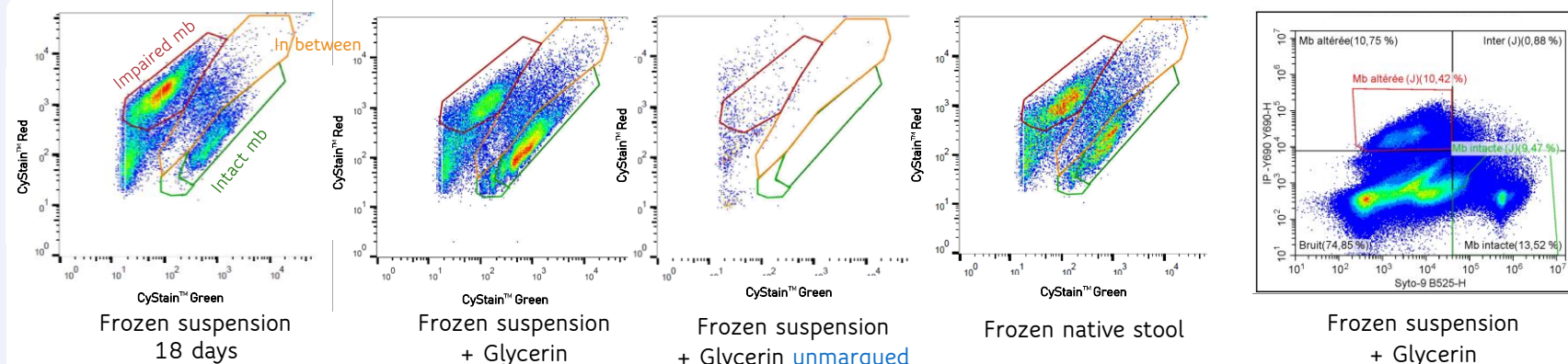
CytoFLEX® S (Beckman  
Coulter®)

Staining: SYTO™9/  
Iodure de propidium (IP)

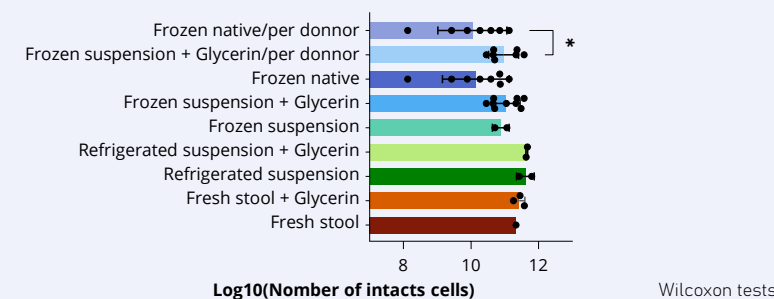
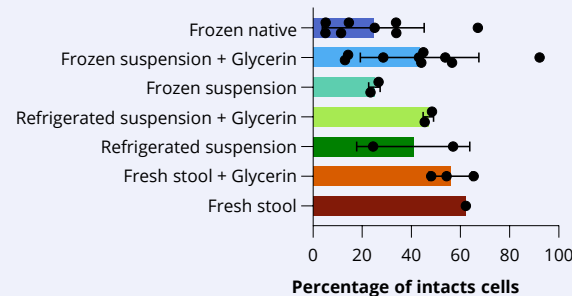
Simplicate, 4 stools

Blue and yellow lasers

## Results



	Cyflow®	CytoFLEX®	Criteria	Results
Price	Equivalent ~ 50 k€		CV permeability of membrane	<13%
Training	Complete by the supplier		CV enumeration	<2% (in log MO/g of stool)
Number of manipulation	+	- (automated homogenization and rinsing)	Enumeration	10 <sup>9</sup> and 10 <sup>12</sup> MO/g of stool, consistent with the literature
Upgrade	Change device	Laser addition possible without changing the device	Stool preparation	<ul style="list-style-type: none"> <li>No prior centrifugation or filtration</li> <li>Dilution to 10<sup>-6</sup>: to achieve 10<sup>9</sup>-10<sup>11</sup> MO/g of stool (if outside this range, analyze the lower or higher dilution)</li> </ul>
Ergonomics	Integrated computer	Integrated sheath fluid and waste bottles		



## Conclusion

- ⇒ Rapid, repeatable results
- ⇒ Method does **not** provide information on: type of MO or other components of the fecal microbiota
- ⇒ Collaboration with other teams to enhance expertise
- ⇒ Supports innovation towards new pharmaceutical forms